



## PRODUCT SPECIFICATION SHEET

### Cetrimide Agar Plate (RP002H)

#### Intended Use

Medium for selective isolation and subculture of *Clostridium sporogenes* in compliance with the harmonized method of USP/EP/BP/JP.

#### Product Summary and Explanation

King et al.<sup>(1)</sup> developed Medium A (Tech Agar) for the enhancement of pyocyanin production by *Pseudomonas*. Cetrimide Agar has the formula for Tech Agar but is modified by the addition of cetrimide (cetyl trimethyl ammonium bromide) for the selective inhibition of organisms other than *P. aeruginosa*. In 1951, Lowburry first reported the use of cetrimide as an agent for selective isolation of *Pseudomonas*.<sup>(2)</sup> The concentration was later reduced, because of the increased purity of the inhibitory agent, as reported by Lowbury and Collins in 1955.<sup>(3)</sup> Brown and Lowbury employed incubation at 37°C with examination after 18 and 42 hours of incubation.<sup>(4)</sup> Strains of *P. aeruginosa* are identified from specimens by their production of pyocyanin, a blue, water-soluble, nonfluorescent, phenazine pigment in addition to their colonial morphology<sup>(5)</sup> and the characteristic grapelike odor of aminoacetophenone.<sup>(6)</sup> *P. aeruginosa* is the only species of *Pseudomonas* or gram-negative rod known to excrete pyocyanin. Cetrimide Agar, therefore, is a valuable culture medium in the identification of this organism.

This media formulation is in accordance with the harmonized method of USP/EP/BP/JP/IP.<sup>(7-11)</sup> It is used as a selective medium for the isolation of *Pseudomonas aeruginosa* from pharmaceutical products. Cetrimide Agar is widely recommended for use in the examination of cosmetics,<sup>(12)</sup> clinical specimens<sup>(5,13)</sup> for the presence of *P. aeruginosa*, as well as for evaluating the efficacy of disinfectants against this organism.<sup>(14)</sup>

#### Principles of the Procedure

Cetrimide Agar contains pancreatic digest of gelatine which supplies the nutrients necessary to support growth, while glycerin serves as slow and continuous carbon source for the growing cell. Magnesium chloride and potassium sulphate incorporated in the medium enhances the production of pigment pyocyanin, which is a blue-green pigment, diffusing into the medium. This improves detection of *Pseudomonas* on this medium. Presence of magnesium ions can also neutralize EDTA, if present in the sample. Cetrimide (N-acetyl-N,N,N-trimethylammonium bromide) is a quaternary ammonium, cationic detergent compound, which is inhibitory to a wide variety of bacterial species including *Pseudomonas* species other than *P. aeruginosa*. *Pseudomonas aeruginosa* colonies may appear pigmented greenish (under UV light also). Addition of nalidixic acid can aid in inhibiting the growth of accompanying flora.

#### Formula / Liter

Ingredients	Gms / Liter
Pancreatic digest of gelatin	20.00
Magnesium chloride	1.40
Potassium sulphate	10.00
Cetrimide	0.30
Agar	15.00
Final pH: 7.2 ± 0.2 at 25°C	
Formula may be adjusted and/or supplemented as required to meet performance specifications	

#### Precautions

1. Prepared plated media are For *in vitro* Diagnostic Use or For Laboratory Use as labeled.
2. Directions for use should be read and followed carefully.
3. If excessive moisture is observed, invert the bottom over an off-set lid and allow to air dry in order to prevent formation of a seal between the top and bottom of the plate during incubation.





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4. Observe aseptic techniques and established precautions against microbiological hazards throughout all procedures, since it must be assumed that all specimens/samples collected might contain infectious microorganisms.

### Product Deterioration

Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

### Quality Control Specifications

Appearance	Sterile Cetrimide Agar in 90mm plates
Colour	Light amber coloured medium
Reaction	pH : 7.2 ± 0.2 at 25°C
Quantity of medium	25ml of medium in 90mm plates

**Sterility Check:** Passes release criteria.

Cultural response	Cultural characteristics observed after incubation at 30-35 °C for 18-72 hours. Recovery rate is considered as 100% for bacteria growth on Soyabean Casein Digest Agar.
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### Expected Cultural Response:

Sr. No.	Organisms	Results to be achieved				
		Inoculum (CFU)	Growth	Observed Lot value (CFU)	Recovery	Incubation Temperature & Time
1.	<i>Pseudomonas aeruginosa</i> ATCC 9027	50 - 100	good-luxuriant	25 -100	≥50 %	30-35°C ≤18 hrs
2.	<i>Escherichia coli</i> ATCC 8739	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
3.	<i>Pseudomonas aeruginosa</i> ATCC 27853	50 - 100	good-luxuriant	25 - 100	≥50 %	30-35°C 18-24 hrs
4.	<i>Pseudomonas aeruginosa</i> ATCC 25668	50 - 100	good-luxuriant	25 - 100	≥50 %	30-35°C 18-24 hrs
5.	<i>Stenotrophomonas maltophilia</i> ATCC 13637	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
6.	<i>Escherichia coli</i> ATCC 25922	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
7.	<i>Escherichia coli</i> NCTC 9002	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
8.	<i>Staphylococcus aureus</i> ATCC 6538	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
9.	<i>Staphylococcus aureus</i> ATCC 25923	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
10.	<i>Salmonella typhimurium</i> ATCC 14028	≥10 <sup>3</sup>	inhibited	0	0%	30-35°C ≥72 hrs
11.	<i>Proteus mirabilis</i> ATCC 29906	≥10 <sup>3</sup>	inhibited	0	0%	30-35 °C ≥72 hrs

The organisms listed are the minimum that should be used for quality control testing.





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### Test Procedure

1. Refer to appropriate references for standard test procedures. For the isolation of *Pseudomonas aeruginosa*, plates of Cetrimide Agar should be inoculated from non-selective medium such as Soybean Casein Digest Medium (DM277H). If the count is high the test sample can be directly inoculated onto this medium.

### Results

1. Colonies that are surrounded by a blue-green pigment and fluoresce under short wavelength (254 nm) ultraviolet light may be presumptively identified as *Pseudomonas aeruginosa*. Note, however, that certain strains of *P. aeruginosa* may not produce pyocyanin.
2. Other species of *Pseudomonas* do not produce pyocyanin, but fluoresce under UV light.
3. Most non-*Pseudomonas* species are inhibited, and some species of *Pseudomonas* may also be inhibited.
4. Gram staining, biochemical tests and serological procedures should be performed to confirm findings.

### Storage

On receipt, store plates at 20-25°C.

### Expiration

Refer to the expiration date stamped on the pack. Prepared plates stored in their original sleeve wrapping at 20-25°C until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times.

### Product Disposal

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

### Limitations of the Procedure

1. The type of peptone used in the base may affect pigment production.
2. No single medium can be depended upon to exhibit all pigment-producing *P. aeruginosa* strains.
3. Occasionally some enterics will exhibit a slight yellowing of the medium; however, this coloration is easily distinguished from fluorescein production since this yellowing does not fluoresce.
4. Some nonfermenters and some aerobic sporeformers may exhibit a water-soluble tan to brown pigmentation on this medium. *Serratia* strains may exhibit a pink pigmentation.
5. Studies of Lowbury and Collins showed *P. aeruginosa* may lose its fluorescence under UV light if the cultures are left at room temperature for a short time. Fluorescence reappears when plates are reincubated.
6. Consult appropriate texts for detailed information and recommended procedures.
7. .

### Packaging

**Product Name : Cetrimide Agar Plate**

**Product Code : RPO02H**

**Available Pack sizes : Pack of 10 plates**

### References

1. King, Ward, and Raney. 1954. J. Lab. Clin. Med. 44 :301.
2. Lowbury. 1951. J. Clin. Pathol. 4 :66.
3. Lowbury and Collins. 1955. J. Clin. Pathol. 8 :47.
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5. Blondel-Hill, Henry and Speert. 2007. In Murray, Baron, Jorgensen, Landry and Pfaller (eds.), Manual of clinical microbiology, 9th ed. American Society for Microbiology, Washington, D.C.
6. Gilardi. 1991. In Balows, Hausler, Herrmann, Isenberg and Shadomy (eds.), Manual of clinical microbiology, 5th ed. American Society for Microbiology, Washington, D.C.
7. The United States Pharmacopoeia, 2011 The United States Pharmacopoeial Convention. Rockville, MD.
8. British Pharmacopoeia, 2011, The Stationery office British Pharmacopoeia.
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- 10. Japanese Pharmacopoeia, 2008.
- 11. Indian Pharmacopoeia, 2010, Govt. of India, Ministry of Health and Family Welfare, New Delhi.
- 12. Hitchens, Tran, and McCarron. 2001. In FDA bacteriological analytical manual online, 8th ed. <http://www.cfsan.fda.gov/~ebam/bam-23.html>.
- 13. Forbes, Sahn, and Weissfeld. 2007. Bailey & Scott's diagnostic microbiology, 12th ed. Mosby Elsevier, St. Louis, Mo.
- 14. Horwitz, (ed). 2002. AOAC Official Method 955.13. Official methods of analysis of AOAC International, 17<sup>th</sup> ed, vol. 1, Rev. 1. AOAC International, Gaithersburg, Md.

### Further Information

For further information please contact your local MICROMASTER Representative.



**MICROMASTER LABORATORIES PRIVATE LIMITED** RP002HPSS, Rev.00, Ver.00/01.02.2016  
 Unit 38/39, Kalpataru Industrial Estate,  
 Off G.B. Road, Near 'R-Mall', Thane (W) - 400607. M.S. INDIA.  
 Ph: +91-22-25895505, 4760, 4681. Cell: 9320126789.  
 Email: [micromaster@micromasterlab.com](mailto:micromaster@micromasterlab.com)

Prepared By	Checked By	Approved By
 01.01.2018	 01.01.2018	 01.01.2018
<b>Microbiologist</b>	<b>Head Quality Control</b>	<b>Head Quality Assurance</b>

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