



PRODUCT SPECIFICATION SHEET

Antibiotic Assay Medium F (DM752B)

Intended Use

Antibiotic Assay Medium F (DM752B) is recommended for microbiological assay of Nystatin using *Saccharomyces cerevisiae* in compliance with BP.

Product Summary and Explanation

Antibiotic assay media are identified numerically with names assigned by Grove and Randall in *Assay Methods of Antibiotics*.⁽¹⁾ The activity (potency) of an antibiotic can be demonstrated under suitable conditions by its inhibitory effect on microorganisms.⁽²⁾ An assay is made to determine the ability of an antibiotic to kill or inhibit the growth of living microorganisms. Biological tests offer the most convenient means of performing an assay,⁽³⁾ since a reduction in the antimicrobial activity of a specific antibiotic reveals changes not usually displayed by chemical methods.⁽⁴⁾ Antibiotic assays are performed by the cylinder plate method and the turbidimetric "tube" assay. The cylinder plate method, first described by Abraham et al.⁽⁵⁾ for the assay of penicillin, was later modified by Foster and Woodruff⁽⁶⁾ and by Schmidt and Moyer.⁽⁷⁾ The choice of methodology is often based on many factors, including relative ease of performance, flexibility and use of automated or semi-automated devices for both identification and susceptibility testing.⁽⁸⁾ Antibiotic assay Medium F is recommended for the microbiological assay of Nystatin and Amphotericin B using *Saccharomyces cerevisiae* and *Candida tropicalis*. This medium is formulated in accordance with the British Pharmacopoeia.⁽⁹⁾

Principles of the Procedure

Antibiotic Assay Medium F contains combination of peptone, yeast extract and beef extract which provides nitrogenous, vitamins, amino acids and other essential growth nutrients. Glucose monohydrate in the medium provides enhanced source of carbon and energy. Sodium chloride maintains osmotic equilibrium in the medium which helps the cell viability and integrity. Higher agar concentration provides solid substratum for growth of colonies and controls the diffusion of antibiotics.

Cylinder Plate Assay

This method is based on the diffusion of an antibiotic solution from a cylinder placed on the surface of an inoculated agar medium. After incubation the diameter of a zone of inhibition depends, in part, on the concentration or activity of the antibiotic. This method is used in the assay of commercial preparations of antibiotics, as well as in the quantitative determination of antibiotics in body fluids, animal feeds and other materials.

Prediffusion of antibiotics for 10-20 mins in the agar by incubating at temperature below the optimal growth temperature for microorganism would facilitate better diffusion of antibiotics followed by incubation of plates for microbial growth.

Note : Recommended for the microbiological assay of Amphotericin B and Nystatin.

Formula / Liter

Ingredients	Gms / Liter
Peptone	9.40
Yeast extract	4.70
Beef extract	2.40
Sodium chloride	10.00
Glucose monohydrate	10.00
Agar	23.50
Final pH: *6.0 ± 0.1 at 25°C	
Formula may be adjusted and/or supplemented as required to meet performance specifications	
*While assaying Amphotericin B adjust the pH to 6.1±0.1	



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Precautions

1. For Laboratory Use only.
2. IRRITANT. Irritating to eyes, respiratory system, and skin.
3. Freshly prepared plates should be used for antibiotic assays.
4. All conditions in the microbiological assay must be controlled carefully.
5. The use of standard culture medium in the test is one of the important steps for obtaining good results.

Directions

1. Suspend 59.09 grams in one litre of R-water/ purified /distilled water.
2. Heat to boiling to dissolve the medium completely.
3. Dispense and autoclave at 121°C, 15 psi pressure, for 15 minutes / validated cycle.
4. Adjust the pH of the medium, using freshly prepared buffer solution as recommended by the British Pharmacopoeia for the antibiotic assayed.

Quality Control Specifications

Dehydrated Appearance	Cream to yellow coloured homogeneous free flowing powder
Prepared Medium	Light yellow coloured clear to slightly opalescent gel forms in Petri plates
Reaction of 5.91% solution	pH : 6.0 ± 0.1 at 25°C
Gel Strength	Firm, comparable with 2.35% Agar gel

Expected Cultural Response: Cultural characteristics observed after an incubation at specified temperature for 18-24 hours. (*-While assaying Amphotericin B, adjust the pH to 6.0-6.20)

Sr. No.	Organisms	Results to be achieved				Incubation Temperature
		Inoculum (CFU)	Growth	Recovery	Antibiotic Assayed	
1.	<i>Saccharomyces cerevisiae</i> ATCC 9763	50 - 100	good-luxuriant	≥70%	Amphotericin B, Nystatin	35-37°C 30-32°C
2.	<i>Candida albicans</i> CIP1433-83	50 - 100	good-luxuriant	≥70%	Nystatin	30-37°C

The organisms listed are the minimum that should be used for quality control testing.

Test Procedure

Preparation of Stock cultures

1. Maintain stock cultures on agar slants and make transfers at 1- or 2-week intervals.
2. Using sterile purified water, saline or Antibiotic Medium No. 3, prepare the inoculum for assay by washing growth from a fresh 24-48 hour agar slant and further dilute the culture to obtain the desired organism concentration.

Cylinder Plate Assay

1. Use 20 × 100 mm glass or plastic Petri dishes with sufficient depth so that cylinders used in the assay will not be pushed into the medium by the cover.
2. Use stainless steel or porcelain assay cylinders having the following dimensions (± 0.1 mm): 8 mm outside diameter, 6 mm inside diameter and 10 mm long. Clean the cylinders carefully to remove all residues, using an occasional acid bath (i.e., with approximately 2N nitric acid or with chromic acid).
3. Four or six cylinders are generally used per plate, evenly spaced on a 2.8 cm radius.
4. For assuring accurate assays, use a level surface for working to obtain uniformly thick base and seed layers in the Petri dish.
5. Allow the base layer to solidify and then overlay the seed layer containing a proper concentration of the test organism. The amount of medium in the layers varies for different antibiotics, with most assays specifying a 21 ml base layer and a 4 ml seed layer.
6. In any case, dishes with flat bottoms are required to assure complete coverage of the bottom of the dish when



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	Checked By	Approved By
 01.01.2018	 01.01.2018	 01.01.2018
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