

PRODUCT SPECIFICATION SHEET

Antibiotic Assay Medium No. 19 (DM025)

Intended Use

Antibiotic Assay Medium No. 19 (DM025) is recommended for microbiological assay of Amphotericin B, Candidin and Nystatin using *Saccharomyces cerevisiae*.

Product Summary and Explanation

Antibiotic assay media are prepared according to the specifications of the USP⁽¹⁾, European Pharmacopeia⁽²⁾ and AOAC International.⁽³⁾ The antibiotic media are identified numerically with names assigned by Grove and Randall in *Assay Methods of Antibiotics*.⁽⁴⁾ The activity (potency) of an antibiotic can be demonstrated under suitable conditions by its inhibitory effect on microorganisms.⁽¹⁾ Reduction in antimicrobial activity may reveal changes not demonstrated by chemical methods.⁽¹⁾ Antibiotic assays are performed by the cylinder plate method and the turbidimetric "tube" assay. The cylinder plate method, first described by Abraham et al.⁽⁵⁾ for the assay of penicillin, was later modified by Foster and Woodruff⁽⁶⁾ and by Schmidt and Moyer.⁽⁷⁾

Antibiotic Assay Medium No. 19 corresponds to the use described in *Outline of Details for Official Microbiological Assays of Antibiotics* and is as per specification of Krishbaum and Arett.⁽⁸⁾ This medium used as seed agar for assay of antifungal agents like Amphotericin B, Nystatin, Netamycin and Candidin etc. This medium can also be used for maintenance and inoculum development of *Saccharomyces cerevisiae* as well as for assaying mycostatic activity in pharmaceutical formulations.

Principles of the Procedure

Antibiotic Assay Medium No. 19 contains peptic digest of animal tissue, yeast and beef extract which supplements essential nutrients, minerals and growth factors for the growth of test organism. Dextrose serves as an enhanced source of carbon and energy. Sodium chloride maintains the osmotic equilibrium of the medium which helps to maintain the cell integrity and viability.

Cylinder Plate Assay

This method is based on the diffusion of an antibiotic solution from a cylinder placed on the surface of an inoculated agar medium. After incubation, the diameter of a zone of inhibition is observed which depends, in part, on the concentration or activity of the antibiotic. The results depend on critical rates of diffusion of the antibiotic, critical growth rates of the standard organisms and critical minimal inhibitory coefficient levels of each organism. This method is used in the assay of commercial preparations of antibiotics, as well as in the quantitative determination of antibiotics in body fluids, animal feeds and other materials.

Pre-diffusion of microorganism for 10-20 minutes in the agar by incubating at temperature below the optimal growth temperature for microorganism would facilitate better diffusion of antibiotics followed by incubation of plates for microbial growth.

Note : Recommended in the microbiological assay of Amphotericin B, Candidin and Nystatin

Formula / Liter

Ingredients	Gms / Liter
Peptic digest of animal tissue (Peptone)	9.40
Yeast extract	4.70
Beef extract	2.40
Dextrose	10.00
Sodium chloride	10.00
Agar	23.50
Final pH: 6.1 ± 0.2 at 25°C	
Formula may be adjusted and/or supplemented as required to meet performance specifications	

Precautions



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1. For Laboratory Use only.
2. IRRITANT. Irritating to eyes, respiratory system, and skin.
3. Freshly prepared plates should be used for antibiotic assays.
4. All conditions in the microbiological assay must be controlled carefully.
5. The use of standard culture medium in the test is one of the important steps for obtaining good results.

Directions

1. Suspend 60 grams of the medium in one liter of distilled water.
2. Heat to boiling, to dissolve the medium completely.
3. Autoclave at 121°C, 15 psi pressure, for 15 minutes / validated cycle.

Quality Control Specification

Dehydrated Appearance	Cream to yellow homogeneous free flowing powder
Prepared Medium	Yellow coloured, clear to slightly opalescent gel forms in Petri plates
Reaction of 6.0% solution	pH 6.1 ± 0.2
Gel Strength	Firm, comparable with 2.35% Agar gel

Expected Cultural Response: Cultural characteristics observed after an incubation at 29-31°C for 24-48 hours.

Sr. No.	Organisms	Results to be achieved			
		Inoculum (CFU)	Growth	Recovery	Antibiotics Assayed
1.	<i>Saccharomyces cerevisiae</i> ATCC 2601	50-100	good-luxuriant	≥70%	Nystatin
2.	<i>Saccharomyces cerevisiae</i> ATCC 9763	50-100	good-luxuriant	≥70%	Amphotericin B, Candicidin

The organisms listed are the minimum that should be used for quality control testing.

Test Procedure

Preparation of Stock cultures

1. Maintain stock cultures on agar slants and make transfers at 1- or 2-week intervals.
2. Using sterile purified water, saline or Antibiotic Medium 3, prepare the inoculum for assay by washing growth from a fresh 24-48 hour agar slant and further dilute the culture to obtain the desired organism concentration.
3. In some turbidimetric assays, an 18- 24hour culture of the test organism grown in Antibiotic Medium 3, diluted to obtain the optimal number of organisms, is used.

Cylinder Plate Assay

1. Use 20 × 100 mm glass or plastic Petri dishes with sufficient depth so that cylinders used in the assay will not be pushed into the medium by the cover.
2. Use stainless steel or porcelain assay cylinders having the following dimensions (±0.1 mm): 8 mm outside diameter, 6 mm inside diameter and 10 mm long. Clean the cylinders carefully to remove all residues, using an occasional acid bath (i.e., with approximately 2N nitric acid or with chromic acid).
3. Four or six cylinders are generally used per plate, evenly spaced on a 2.8 cm radius.
4. For assuring accurate assays, use a level surface for working to obtain uniformly thick base and seed layers in the Petri dish.
5. Allow the base layer to solidify and then overlay the seed layer containing a proper concentration of the test organism. The amount of medium in the layers varies for different antibiotics, with most assays specifying a 21 mL base layer and a 4 mL seed layer.
6. In any case, dishes with flat bottoms are required to assure complete coverage of the bottom of the dish when small amounts of base medium are used. Tilt the plate to obtain even coverage of the base layer by the seed layer and allow it to solidify in a level position. Plates should be used the same day as prepared.

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Results

Refer to appropriate references and specific test procedures.

Storage

Store the sealed bottle containing the dehydrated medium at 10 - 30°C. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light.

Expiration

Refer to the expiration date stamped on the container. The dehydrated medium should be discarded if not free flowing, or if the appearance has changed from the original color. Expiry applies to medium in its intact container when stored as directed.

Limitations of the Procedure

For a complete discussion of antibiotic assay methods, refer to appropriate procedures outlined in the references.^{1,2,3}

Packaging

Product Name : Antibiotic Assay Medium No.19

Product Code : DM025

Available Pack sizes : 500gm

References

1. United States Pharmacopeial Convention, Inc. 2008. The United States pharmacopeia 31/ The national formulary 26, Supp. 1, 8-1-08, online. United States Pharmacopeial Convention, Inc., Rockville, Md.
2. Council of Europe. 2002. European pharmacopeia, 4th ed. Council of Europe, Strasbourg, France.
3. Horwitz (ed.). 2007. Official methods of analysis of AOAC International, 18th ed., online. AOAC International, Gaithersburg, Md.
4. Grove and Randall. 1955. Assay methods of antibiotics. Medical Encyclopedia, Inc. New York, N.Y.
5. Abraham, Chain, Fletcher, Florey, Gardner, Heatley and Jennings. 1941. *Lancet* ii: 177.
6. Foster and Woodruff. 1943. *J. Bacteriol.* 46:187.
7. Schmidt and Moyer. 1944. *J. Bacteriol.* 47:199.
8. Kirshbaum and Arret. 1967. *J. Pharm. Sci.* 56:512.

Further Information

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