

# Baird Parker Agar Base (DM1031)

### Intended Use

Baird Parker Agar Base (DM1031) is recommended for isolation and enumeration of coagulase positive Staphylococci from foods and another specimens using FPT Inhibitor Supplement.

#### Product Summary and Explanation

Baird Parker Agar was developed by Baird-Parker <sup>(1,2)</sup> from the Tellurite-glycine formulation of Zebovitz et al<sup>(3)</sup> for isolation of S. aureus from foods. Baird Parker medium was reported to be the best medium for selective detection of coagulase positive and entero-toxigenic Staphylococcus. <sup>(6)</sup> Researchers have found a high correlation between the coagulase test and the presence of clear zone of lypolysis in this medium, which is because of the lecithinase of Staphylococci that breakdown, the egg yolk. On the other hand, some studies show that almost 100% of coagulase positive Staphylococci are capable of reducing tellurite, which produces black colonies, whereas other Staphylococci cannot reduce tellurite. Baird Parker Agar was found to be less inhibitory to Staphylococcus aureus than other media at the same time being more selective. <sup>(4, 5, 6)</sup> Subsequently the use of Baird-Parker Agar was officially adopted by AOAC International <sup>(7)</sup> and is recommended in the USP for use in the performance of Microbial Limit Tests. <sup>(8)</sup> Recently, ISO committee has also recommended this medium for the isolation and enumeration of Staphylococci.

The identity of Staphylococcus aureus isolated on Baird-Parker Agar must be confirmed with a coagulase reaction. Baird- Parker Agar base is different from Baird- Parker Agar medium can also be used to detect coagulase activity by adding fibrinogen plasma, rabbit plasma and trypsin inhibitor to prevent fibrinolysis.

#### Principles of the Procedure

In Baird Parker Agar Base beef extract, yeast extract and casein enzymic hydrolysate provides essential mineral, vitamin, nitrogenous compounds and other growth requirements. Sodium pyruvate is incorporated to protect injured cells and help recovery and growth of *S. aureus* without destroying the selectivity. Lithium chloride and potassium tellurite inhibit most of contaminating microflora except *S. aureus*. The tellurite additive is toxic to egg yolk- clearing strains other than *S. aureus*. The reduction of tellurite is a characteristic of coagulase-positive staphylococci, and imparts a black color to the colonies. Glycine, pyruvate enhances growth of *Staphylococcus*.

Formula / Liter									
Ingredients	Gms / Liter								
Pancreatic digest of Casein	10.00								
Beef extract	5.00								
Yeast extract	1.00								
Glycine	12.00								
Sodium pyruvate	10.00								
Lithium chloride	5.00								
Agar	20.00								
Final pH: 7.0 ± 0.2 at 25°C									
Formula may be adjusted and/or supplemented specifications	as required to meet performance								

#### **Precautions** checked

- 1. For Laboratory Use only.
- 2. IRRITANT. Irritating to eyes, respiratory system, and skin.
- 3. Lithium Chloride is harmful. Avoid all bodily contact and inhalation of vapours. On contact with skin wash with plenty of water immediately.

#### Directions

- 1. Suspend 6.3 grams in 90 ml purified / distilled water.
- 2. Heat to boiling to dissolve the medium completely.
- 3. Sterilize by autoclaving at 15 lbs. pressure (121°C) for 15 minutes.





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4. Cool to 45-50°C and aseptically add rehydrated content of 1 vial of FPT Inhibitor Supplement (FD195).

5. Mix well and pour into sterile Petri plates.

# **Quality Control Specifications**

Quality control opecifications						
Dehydrated Appearance	2hydrated Appearance Cream to yellow homogeneous free flowing powder					
Prepared Medium	Basal medium: Yellow coloured clear to slightly opalescent gel, After addition of of					
	Fibrinogen plasma trypsin inhibitor supplement (MS118): Yellow coloured opaque gel					
	forms in Petri plates.					
Reaction of 6.3% Solution	pH : 7.0 ± 0.2 at 25°C					
Gel Strength	Firm, comparable with 1.5% Agar gel					

**Expected Cultural Response:** Cultural characteristics observed with added FPT Inhibitor Supplement (MS118), after an incubation at 35-37°C for 24-48 hours

	Organisms	Results to be achieved					
Sr. No.		Inoculum (CFU)	Growth	Lot value (CFU)	Recovery	Colour of colony	Lecithinase
1.	Staphylococcus aureus ATCC 6538	50 -100	good- luxuriant	25-100	>=50 %	grey-black shiny	Positive, opaque zone around the colony
2.	Staphylococcus aureus ATCC 25923	50 -100	good- luxuriant	25-100	>=50 %	grey-black shiny	Positive, opaque zone around the colony
3.	Proteus mirabilis ATCC 25933	50 -100	good- luxuriant	50-100	<b>≻</b> =50 %	brown - black	Negative
4.	Micrococcus luteus ATCC 10240	50-100	poor- good	15 -40	30-40%	shades of brown- black (very small)	Negative
5.	Staphylococcus epidermidis ATCC 12228	50-100	poor- good	15 -40	30-40%	black	Negative
6.	Bacillus subtilis ATCC 6633	50-100	none- poor	0 -10	0 -10	dark brown matt	Negative
7.	Escherichia coli ATCC 25922	50-100	none- poor	0 -10	0 -10	large brown black	Negative

The organisms listed are the minimum that should be used for quality control testing.

### Test Procedure

- 1. Food samples are macerated in suitable broth medium, diluted as desired and the dilutions spread-inoculated onto the agar surfaces, which should be dry when inoculated.
- 2. Alternatively, inoculate the material to be examined (0.1 ml per plate of diameter 90-100 mm),
- 3. Incubate plates aerobically for 24 hours at 35 37°C.
- 4. Perform the coagulase test on the colonies with the typical characteristics, which have developed during the incubation period.
- 5. Plates should be used on the same day of preparation or within 48 hours, to avoid the loss of definition in the precipitated zones.
- 6. To detect coagulase activity by adding Fibrinogen Plasma Trypsin Inhibitor supplement (MS118), is dissolved in 10 ml sterile distilled water added to 90 ml sterile molten media kept at 45-50°C. On this medium coagulase positive colonies appear white to grey-black surrounded by an opaque zone due to coagulase activity within 24-48 hours



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incubation at 35°C. Reduction in tellurite is necessary because of absence of egg yolk emulsion. This results in translucent agar and white to grey coloured colonies of Staphylococci. For quantitative results select 20-200 colonies. Count Staphylococcus aureus like colonies and test them for coagulase reaction. Report Staphylococcus aureus per gram of food.

7. Refer appropriate references for detailed instructions of specific procedures.

# Results

- 1. Coagulase-positive typical colonies of *Staphylococcus aureus* produce grey-black, shiny, convex colonies with entire margins and clear zones with or without an opaque zone.
- 2. Coagulase-negative staphylococci generally produce poor or no growth. If growth occurs, colonies are black; typical clear or opaque zones are rare.

### Storage

Store the sealed bottle containing the dehydrated medium at 10 - 30°C. Once opened and recapped, place container in a low humidity environment at the same storage temperature. Protect from moisture and light.

### Expiration

Refer to the expiration date stamped on the container. The dehydrated medium should be discarded if not free flowing, or if the appearance has changed from the original color. Expiry applies to medium in its intact container when stored as directed.

## Limitations of the Procedure

- 1. Baird-Parker Agar is selective for coagulase-positive staphylococci, but other bacteria may grow.
- 2. Microscopic examination and biochemical tests will differentiate coagulase-positive staphylococci from other organisms.
- 3. Consult appropriate texts for detailed information and recommended procedures.

### Packaging

Product Name : Baird Parker Agar Base Product Code : DM1031 Available Pack sizes : 100gm / 500gm

### References

- 1. Baird-Parker, A.C. 1962, J.Appl.Bact., 25: 12.
- 2. Baird-Parker, A.C. and Davenport, E., 1965, J.Appl. Bact., 28: 390.
- 3. Zebovitz, E., Evans J.B. & Niven C.F., (1955), J. Bact; 70:686.
- 4. Tardio and Baer, 1971, J. Assoc. Off. Anal. Chem., 54:728.
- 5. Baer, 1971, J. Assoc. Off. Anal. Chem., 54:732.
- 6. Assoc. off. Anal. Chem., 1971, 54:401.
- 7. Horwitz (Ed.), 2000, Official methods of analysis of AOAC International, 17th Ed., Vol. I., AOAC International, Gaithersburg, MD.
- 8. The United States Pharmacopoeia, 2008, USP31, The United States Pharmacopoeial Convention. Rockville, MD.
- 9. International Organization for Standardization (ISO), 1983, Draft ISO/DIS 6888.
- 10. Smith B. A. and Baird-Parker A.C., 1964, J. Appl. Bacter

### Further Information

For further information please contact your local MICROMASTER Representative.



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