# PRODUCT SPECIFICATION SHEET

## Soyabean Casein Digest Agar Plate contact plate,Lockable (- irradiated) (Triple Pack) (CP001IT)

**Intended Use**

Soyabean Casein Digest Agar Plate contact plate (- irradiated) (CP001IT) is recommended for cultivation of wide variety of microorganisms and for Sterility testing in pharmaceuticals.

**Product Summary and Explanation**

Soyabean Casein Digest Agar Plate w/Lecithin and Polysorbate may be employed to establish and monitor cleaning techniques and schedules.[1-4] Collection of "samples" from identical areas before and after treatment with disinfectant yields data useful in evaluating cleaning procedures in environmental sanitation. Tryptone Soyabean Casein Digest Agar Plate w/Lecithin and Polysorbate is recommended for water-miscible cosmetic products containing preservatives.[5] This medium is used in RODAC (Replicate Organism Detection and Counting) plates[6] for the detection and enumeration of microorganisms present on surfaces of sanitary importances.[7,8] The presence and number of microorganisms on a flat impervious surface is determined by the appearance of colonies on the surface of the medium following application to the test surface and incubation.[9,10]

The RODAC plate has a marked grid to facilitate counting organisms. The RODAC SL (Secure Lid) has three lugs on the base, providing a tight fit between lid and base to reduce accidental contamination. The 100 × 15 mm and the 150 × 15 mm style plates can be used for active and passive air sampling. These plates are also designed for personnel monitoring of finger tips. After counting the colonies, carry out biochemical testing for identification.[11]

**Principles of the Procedure**

Soyabean Casein Digest Agar Plate contains pancreatic digest of casein and papaic digest of soyabean which provides amino acids, long chain peptides and essential nutrients required for the growth of microorganisms. Sodium chloride maintains the osmotic balance

**Formula / Liter**

| Ingredients | Gms / Litre |
|---|---|
| Casein enzymic hydrolysate | 15.00 |
| Papaic digest of soyabean meal | 5.00 |
| Sodium chloride | 5.00 |
| Agar | 15.00 |
| Formula may be adjusted and/or supplemented as required to meet performance specifications | |

**Precautions**

1. Prepared plated media are For *in vitro* Diagnostic Use or For Laboratory Use as labeled.
2. Directions for use should be read and followed carefully.
3. If excessive moisture is observed, invert the bottom over an off-set lid and allow to air dry in order to prevent formation of a seal between the top and bottom of the plate during incubation.
4. Observe aseptic techniques and established precautions against microbiological hazards throughout all procedures, since it must be assumed that all specimens/samples collected might contain infectious microorganisms.

**Product Deterioration**

Do not use plates if they show evidence of microbial contamination, discoloration, drying, cracking or other signs of deterioration.

**Directions**

Either streak, inoculate or surface spread the test inoculum (50-100 CFU) aseptically on the plate.

**Quality Control Specifications**

| | |
|---|---|
| **Appearance** | Sterile Tryptone Soya Agar in contact plates (gamma-irradiated) |
| **Colour** | Light yellow coloured medium |
| **Reaction** | 7.10- 7.50 |
| **Quantity of medium** | 18 ml of medium in 55 mm contact plates |

**Dose of irradiation :** 10.00- 25.00

**Sterility Check:** Passes release criteria.

**Expected Cultural Response:** Growth Promotion was observed after an incubation at 30-35°C for 18-24 hours for bacteria and for 20-25°C <=5 days for fungus. Recovery rate is considered 100% for bacteria growth on soya Agar and fungus growth on Sabouraud Dextrose Agar.

| Sr. No. | Organisms | Results to be achieved CFU) | | | | | |
|---|---|---|---|---|---|---|---|
| | | Inoculum (CFU) | Growth | Observed Lot value (CFU) | Recovery | Incubation Tempera-ture | Incubation Period |
| 1. | *Bacillus subtilis ATCC 6633* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 2. | *Staphylococcus aureus ATCC 25923* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 3. | *Staphylococcus aureus ATCC 6538* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 4. | *Escherichia coli ATCC 25922* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 5. | *Escherichia coli ATCC 8739* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 6. | *Escherichia coli NCTC 9002* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 7. | *Pseudomonas aeruginosa ATCC 27853* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 8. | *Pseudomonas aeruginosa ATCC 9027* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 9. | *Salmonella Abony NCTC 6017* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 10. | *Micrococcus luteus ATCC 9341* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 11. | *Streptococcus pneumonia ATCC 6305* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 12. | *Salmonella Typhimurium ATCC 14028* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | 18 -24 hrs |
| 13. | *Candida albicans ATCC 10231* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | <=5 d |
| 14. | *Candida albicans ATCC 2091* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 30 -35 °C | <=5 d |
| 15. | *Aspergillus brasiliensis ATCC 16404* | 50 - 100 | good-luxuriant | 25 -70 | 50-70% | 30 -35 °C | <=5 d |
| 16. | *Aspergillus brasiliensis ATCC 16404* | 50 - 100 | luxuriant | 35 -100 | >=50 % | 20 -25 °C | <=5 d |
| 17. | *Candida albicans ATCC 10231* | 50 - 100 | luxuriant | 25 -100 | >=70 % | 20 -25 °C | <=5 d |
| 18. | *Candida albicans ATCC 2091* | 50 - 100 | luxuriant | 35 -100 | >=70 % | 20 -25 °C | <=5 d |

The organisms listed are the minimum that should be used for quality control testing.

**Test Procedure**
Refer appropriate references for standard test procedures.

**Results**
Refer appropriate references and procedures for interpretation of results. After counting the colonies, carry out biochemical testing for identification.

**Storage**
On receipt, store plates at 15-25°C.

**Expiration**
Refer to the expiration date stamped on the pack. Prepared plates stored in their original sleeve wrapping at 15-25°C until just prior to use may be inoculated up to the expiration date and incubated for recommended incubation times.

**Product Disposal**

After use, prepared plates, specimen/sample containers and other contaminated materials must be sterilized before discarding.

**Limitations of the Procedure**

1. Some diagnostic tests may be performed with the primary plate. However, a pure culture is recommended for the majority of biochemical tests and other identification procedures.
2. Consult appropriate references for further information.

**Packing Information**

It is triple layered packing containing 10 No. of plates. The primary packaging bag contains 5 plates packing with two nos. of a silica gel desiccant bag and it is packed by using five layered plastic bag.
Then these two 5 plates stacks it is packed into second plastic bag which is labeled and then 10plates into tertiary packing (Plastic bag).
Cartons are used for packaging the bagged stacks. The sealing seams of the bags are heat-sealed. The bags allow easy opening without the use of sharp objects such as scissors or knives. Bags can be peeled open at the ends of the stacks by tearing apart both plastic films of the bag. Apply aseptic techniques. Once the outer bag is opened, appropriate measures should be used to maintain the sterility of the inner bags and the contents.

**Packaging**
**Product Name : Soyabean Casein Digest Agar Plate contact plate**
**Product Code : CP001IT**
**Available Pack sizes : □ -irradiated, Triple Pack (Pack of 10 plates)**

**References**

1. Vesley and Michaelson. 1964. Health Lab. Sci. 1:107.
2. Pryor and McDuff. 1969. Exec. Housekeeper, March.
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6. Hall and Hartnett. 1964. Public Health Rep. 79:1021.
7. Richardson (Ed)., 1985, Standard Methods for the Examination of Dairy Products, 15th ed., APHA, Washington, D.C.
8. MacFaddin J.F., 1985, Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria, Vol. I, Williams and Wilkins, Baltimore.
9. McGowan. 1985. In Lennette, Balows, Hausler and Shadomy (ed.), Manual of clinical microbiology, 4th ed. American Society for Microbiology, Washington, D.C.
10. Bryan. 1995. In Murray, Baron, Pfaller, Tenover and Yolken (ed.), Manual of clinical microbiology, 6th ed. American Society for Microbiology, Washington, D.C.
11. Brummer, 1976, Appl. Environ. Microbiol., 32:80.

**Further Information**

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